# Chemoselective Alkylation for Diversity-Oriented Synthesis of 1,3,4- Benzotriazepin-2-ones and Pyrrolo[1,2][1,3,4]benzotriazepin-6-ones, Potential Turn Surrogates

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**S** Supporting Information



ABSTRACT: 1,3,4-Benzotriazepin-2-ones garner interest for medicinal applications, in part due to their relationship with benzodiazepinones. Ten 1,3,4-benzotriazepin-2-ones 6 and 19 and six pyrrolo[1,2][1,3,4]benzotriazepin-6-ones 7 and 23 were prepared in four to seven steps and 4−60% overall yields by a divergent strategy from methyl anthranilate employing chemoselective alkylations of common linear and cyclic precursors to diversify three triazepinone ring positions (N1, N3, and C5). X-ray crystallography demonstrated that benzotriazepinone 19g may serve as a  $\gamma$ -turn mimic.

1,4-Benzodiazepin-2-ones 1 are common targets because of their biological properties and medicinal applications, $\frac{1}{2}$  which may be due in part to their potential to mimic peptide γ-turn secondary str[u](#page-3-0)ctures (Figure 1).<sup>2−5</sup> Although their aza counterparts have



Figure 1. Representative benzodiazepin-2-one, benzotriazepin-2-ones, and  $\gamma$ -turn structures.

received relatively less attention, 1,3,4-benzotriazepin-2-ones 2 possess intriguing biological activity. For example, 5-cyclohexyl triazepinones 3 and 4 have, respectively, exhibited activity as a parathyroid hormone-1 receptor antagonist<sup>6</sup> and an orally active cholecystokinin-2  $(CCK_2)$  antagonist (Figure 1).<sup>7</sup> 1,3,4-Benzotriazepin-2-ones have also been [c](#page-3-0)laimed to possess psychostimulant, antidepressant, anorexigenic, and a[nt](#page-3-0)ihypertensive properties.<sup>8</sup> Although relatively little is known about 1,3,4-triazepin-2-one conformation, similar to the amino acid component in  $1,4$  $1,4$ -diazepin-2-ones, the aza-phenylglycine

residue of 7-chloro-3,5-diphenyl-1,2-dihydro-3H-1,3,4-benzotriazepin-2-one 5 was observed by X-ray crystallography to adopt  $\phi$ - and  $\psi$ -dihedral angle values close to those of the central residue of an ideal  $\gamma$ -turn.<sup>1</sup>

Since original syntheses from 2-aminobenzophenone, $4$  1,3,4benzotriazepin-2-ones [hav](#page-3-0)e been commonly prepared by cyclization of the corresponding hydrazone with a p[ho](#page-3-0)sgene equivalent and by a one-pot annulation with a carbazate often at high temperature (e.g., 190 °C). Ring closure has also been achieved by palladium-catalyzed cyclization of aryl isocyanates and 2-haloaryl hydrazones under microwave irradiation $11$  as well as condensation of anthranilic acid hydrazide with isatins, which provided the corresponding spiro[1,3,4-benzotriaze[pin](#page-3-0)e-2,3′ indole]-2',5(1H,1'H)-diones.<sup>8</sup> Benzotriazepinone skeletons have been alkylated on ring nitrogen and arylated at C5 using copper c[a](#page-3-0)talysis in solution<sup>12</sup> and on microelectrode arrays;<sup>1</sup> however, nitrogen protection has been essential for chemoselectivity.

Diversity-oriented synthesis of 1,3,4-benzotriazepin-2-ones 6 has now been achieved by an approach that avoids nitrogen protection, toxic reagents, and harsh conditions to modify the N1, N3, and C5 positions. Moreover, pyrrolo[1,2][1,3,4] benzotriazepin-6-ones 7 have also been synthesized from common linear precursors prepared from 1-(2-aminophenyl) pent-4-en-1-one 8. This method enhances the utility of amino ketone 8, which has been quantitatively synthesized by a coppercatalyzed cascade addition of vinyl Grignard reagent on methyl anthranilate $14,15$  and used as valuable precursor to make

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<span id="page-1-0"></span>substituted pyrroles  $\boldsymbol{9}^{,14}_\cdot$  quinolines  $\boldsymbol{10}^{,15}_\cdot$  pyrroloquinazolinones 11, <sup>14</sup> 1,4-benzodiazepin-2-ones 12, <sup>5</sup> a[nd](#page-3-0) pyrrolobenzodiazepin2-[one](#page-3-0)s 13 (Figure 2). $<sup>5</sup>$  $<sup>5</sup>$  $<sup>5</sup>$ </sup>



Figure 2. Amino ketone 8 as precursor for heterocycle synthesis.

Amino ketone 8 was acylated with activated carbazate  $14^{16}$  in DCM in the presence of DIEA at room temperature to obtain the key precursor aza-glycinamide 15 in 71% yield on multi[gra](#page-3-0)m scale (Scheme 1). Chemoselective alkylation of the semi-

Scheme 1. Synthesis and Chemoselective Alkylations of Semicarbazone 15 and Ketone 16



carbazone nitrogen of aza-glycinamide 15 was achieved using conditions previously developed for sub-monomer aza-peptide synthesis.<sup>17</sup> Various aza-amino amide analogues 16a−h were thus synthesized from 15 using tetraethylammonium hydroxide as base a[nd](#page-3-0) a diverse set of alkyl halides in THF  $(Table 1)$ .<sup>18</sup>





<sup>a</sup>Et<sub>4</sub>NOH (100 mol %) R<sup>1</sup>X (150 mol %). <sup>b</sup>R = Si(Ph)<sub>2</sub>tBu. <sup>c</sup>Reflux.

Although reactive and primary alkyl halides reacted at room temperature to give 16 in >73% yields, cyclohexylmethyl bromide required heating at reflux to obtain 16g in 52% yield. 1-Bromo-3-chloropropane (150 mol %) reacted with 15 and tetraethylammonium hydroxide (100 mol %) to give azachloropropylglycinamide 16c in 79% yield with minimal amounts of cyclic urea 17c from a second intramolecular alkylation of the aniline nitrogen.

Chemoselective alkylation of the ketone moiety of 16 without reaction on the aniline nitrogen was achieved using LiHMDS (250 mol %) to generate the dianion, which selectively reacted on carbon with various alkyl halides (200 mol %) in THF at 0 °C for 1 h to give branched ketones 18g−m in 35−80% yields (Table 1). Selective alkylation of the ketone enolate may be due to the relative stability and hindered nature of the lithiated urea, which may interact respectively with the neighboring semicarbazone nitrogen and carbonyl oxygen in five and six membered ring chelates. Incomplete alkylation and difficulty in separating product from starting material may account for the lower yields using methyl and ethyl iodides.

1,3,4-Benzotriazepin-2-ones 6a−f were prepared in 32−99% yields from aza-amino amides 16 and 18 by semicarbazone cleavage and cyclization under acidic conditions using 1.0 N aq HCl in THF (Scheme 2, Table 2). Attempts to prepare

# Scheme 2. Cyclization of 16 and 18 and N1-Alkylation of 6



Table 2. 1,3,4-Benzotriazepin-2-ones 6 and 19



benzotriazepinone from unsubstituted semicarbazone 15 were unsuccessful using similar conditions; instead, ions corresponding to oligomer were detected by HPLC−MS analysis of the reaction mixture. Semicarbazone alkylation may favor cyclization by lowering the barrier for urea isomerization to the required Eisomer.<sup>19</sup>  $\alpha$ -Alkyl-branched ketones 18 reacted slower in the cyclization to 6, likely because the neighboring ketone is engaged in a h[ydr](#page-3-0)ogen bond with the aniline NH that disfavors the orientation for nuclophilic attack.<sup>20</sup> The favored hydrogen bonded conformer was illustrated in a comparison of 16h and 18l in deuterium exchange NMR expe[rim](#page-3-0)ents using MeOD- $d_4$  in CDCl3. After 20 h, the amount of exchange of the aniline NH proton with deuterium was >95% for 16h but <20% for 18l under the same conditions. When EtOH was used to competitively hydrogen bond with ketone 18l during the cyclization step, the

reaction time was reduced and the yield increased: 22 h and 53% yield in THF versus 4 h and 63% yield in EtOH.

After cyclization, the aniline nitrogen of triazepinones 6 was chemoselectively alkylated using t-BuOK (120 mol %) and different alkyl bromides (120 mol %) to give the trisubstituted 1,3,4-benzotriazepin-2-ones 19 in 85−100% yields (Table 2). The installment of propargyl, bromoaryl, chloroalkyl, and carboxylate side chains has been demonstrated with [particula](#page-1-0)r interest to further diversify the 1,3,4-benzotriazepin-2-one scaffolds by future employment of such functional groups in orthogonal chemistry, e.g., CuAAC, $^{21}$  cross-coupling, $^{22}$  nucleophilic displacement, $23$  and amide bond forming reactions, respectively. Such chemistry is be[ing](#page-3-0) explored pre[sen](#page-3-0)tly and will be reported in d[ue](#page-3-0) time.

Pyrrolo[1,2][1,4]benzodiazepin-6-ones have garnered interest because of their biological and medicinal relevance. For example, pyrrolobenzodiazepinone (S)-20b has exhibited activity as a non-nucleoside HIV-1 reverse transcriptase inhibitor (Figure 3). $^{24}$  To the best of our knowledge, the aza-variant of this



ring system has never been reported. Pyrrolobenzotriazepinones 7 were thus pursued to further demonstrate the utility of the alkene function. Olefins 16 and 18, respectively, were oxidized using Lemieux−Johnson conditions to give aldehydes 22a−d in 76−84% yields.<sup>25</sup> Aldehydes 22 were then treated with 1.0 N aq HCl in THF at 60 °C to affect semicarbazone cleavage and intramolecular [Pa](#page-3-0)al−Knorr condensation. Pyrrolo[1,2][1,3,4] benzotriazepin-6-ones 7 were isolated in 23−67% yields by column chromatography. The sterically hindered semicarbazone 22d gave pyrrole 7e in 30% yield along with a side product in 23% yield having a molecular ion and spectral properties consistent with imine dimer 21 (Figure 3). Chloropyrrole 7c was isolated in 23% yield from 22c using the acidic cyclization conditions and characterized by its four molecular ions corresponding to the bromine and chlorine isotopes and the pyrrole proton singlet in the NMR spectrum. X-ray analysis of 7c confirmed the structural assignment. Alternatively, by conducting the Paal−Knorr reaction in MeOH in the dark, pyrrolobenzotriazepinone 7d was isolated in 52% yield.

To demonstrate the potential to further diversify pyrrolo-  $[1,2][1,3,4]$ benzotriazepin-6-one 7, analogous conditions were employed as described above for the alkylation of triazepinone 6. The aniline nitrogen of pyrrolobenzotriazepinone 7b was thus alkylated with potassium tert-butoxide and 1-bromo-4-chlorobutane in 84% yield (Scheme 3).

Crystals of 1,3,4-benzotriazepin-2-one 19g and the pyrrolo- [1,2][1,3,4]benzotriazepin-6-one 7c were grown by slow diffusion of n-hexane into samples in EtOAc and subjected to X-ray structural analysis (Figure 4). Substitution of the diazepinone amino acid component by an aza-residue in triazepinone CCK antagonists has previously been used to amplify selectivity for the  $CCK_2$  over  $CCK_1$  receptors.<sup>6</sup> Better accommodation by the  $CCK<sub>2</sub>$  receptor of the achiral triazepinone

Scheme 3. Synthesis and Alkylation of Pyrrolo[1,2][1,3,4]benzotriazepin-6-ones





Figure 4. X-ray crystal structures of 19g and 7c.

may be due to the aza-residue exhibiting adaptive chirality<sup>26</sup> or a relatively flat geometry.

Employing X-ray data to probe the differences o[f t](#page-3-0)heir geometry a comparison of  $\phi$ - and  $\psi$ -dihedral angle values has been made using tri- and disubstituted 1,3,4-benzotriazepin-2 ones 19g and 5, benzodiazepinones R- and S-1a, pyrrolobenzodiand triazepinones 20a and 7c, as well as the central residues of ideal normal and inverse  $\gamma$ -turns (Figure 4, Table 3).<sup>27</sup> The addition of an N1 substituent on benzotriazepinone 5 had limited influence on the dihedral angles relative [to](#page-3-0) its disubstituted counterpart 19g. In comparison to the relatively

![](_page_2_Picture_646.jpeg)

![](_page_2_Picture_647.jpeg)

<span id="page-3-0"></span>similar dihedral angles of benzodiazepinone 1a and ideal γ-turns  $(\phi = 75^{\circ} \pm 3$ , and  $\psi = -65^{\circ} \pm 3$ ), benzotriazepinones 5 and 19g deviated more significantly about the  $\psi$  torsion angle ( $\phi$  = 75°  $\pm$ 1, and  $\psi$  = −65° ± 9). Pyrrolobenzotriazepinone 7c exhibited a  $\phi$ torsion angle  $(-75^\circ \pm 3)$  that was more similar to that of an ideal inverse  $\gamma$ -turn than the value in pyrrolobenzodiazepinone R-20a ( $-75^{\circ} \pm 14$ ); however, the  $\psi$ -torsion angle of 7c differed by 40° away from that of an ideal inverse  $\gamma$ -turn, significantly more than that of its pyrrolobenzodiazepinone counterpart 20a ( $65^{\circ} \pm 8$ ). The dihedral angle values of the aza-amino acid residue in 7c appear to be more closely related to that of the  $i + 2$  residue of a type II'  $\beta$ -turn ( $\phi = -80^\circ \pm 8$ , and  $\psi = 0^\circ \pm 27$ ). The aza-amino acid residue nitrogen in the triazepinones adopted nonplanar configurations existing out of the plane formed by its three neighboring atoms by  $\pm 0.361(1)$  Å for benzotriazepinone 19g and  $\pm 0.015(2)$  Å for pyrrolobenzotriazepinone 7c. For comparison, the deviation from planarity of the corresponding  $\alpha$ -carbons of  $(R)$ -1a,  $(S)$ -1a, and 20a were, respectively, +0.497(2),  $-0.500(3)$ , and  $\pm 0.441$  Å and illustrate that triazepinones 19g and 7c are chiral albeit flatter than their diazepinone counterparts.

In conclusion, chemoselective alkylation was key for conception of efficient diversity oriented strategies to make  $1,3,4$ -benzotriazepin-2-one and pyrrolo $[1,2][1,3,4]$ benzotriazepin-6-one analogues. Without nitrogen protection under relatively mild conditions, benzotriazepinones were made in four to seven steps and 4−60% yields from methyl anthranilate. Using groups suitable for further functionalization, a set of scaffolds was synthesized and shown by X-ray analysis to have potential for γ-turn mimicry. Biological activity of triazepinone library members is under investigation and will be reported in due course.

### ■ ASSOCIATED CONTENT

# **6** Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.orglett.5b03046.

X-ray crystallographic data for 19g and 7c (CIF)

Experimental details and spectroscopic characterization for all compounds (PDF)

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#### **Notes**

The authors declare no competing financial interest.

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### ■ REFERENCES

(1) (a) Sternbach, L. H. J. Med. Chem. 1979, 22, 1−7. (b) Horton, D. A.; Bourne, G. T.; Smythe, M. L. Chem. Rev. 2003, 103, 893−930. (c) Costantino, L.; Barlocco, D. Curr. Med. Chem. 2006, 13, 65−85. (d) Spencer, J.; Rathnam, R. P.; Chowdhry, B. Z. Future Med. Chem. 2010, 2, 1441−1449.

(2) Ramanathan, S. K.; Keeler, J.; Lee, H. L.; Reddy, D. S.; Lushington, G.; Aube, J. Org. Lett. 2005, 7, 1059-1062.

(3) Dufour-Gallant, J.; Chatenet, D.; Lubell, W. D. J. Med. Chem. 2015, 58, 4624−4637.

(4) Sulkowski, T. S.; Childress, S. J. J. Med. Chem. 1964, 7, 386.

(5) Dö rr, A. A.; Lubell, W. D. Org. Lett. 2015, 17, 3592−3595.

(6) McDonald, I. M.; Austin, C.; Buck, I. M.; Dunstone, D. J.; Gaffen, J.; Griffin, E.; Harper, E. A.; Hull, R. A. D.; Kalindjian, S. B.; Linney, I. D.; Low, C. M. R.; Patel, D.; Pether, M. J.; Raynor, M.; Roberts, S. P.; Shaxted, M. E.; Spencer, J.; Steel, K. I. M.; Sykes, D. A.; Wright, P. T.; Xun, W. J. Med. Chem. 2007, 50, 4789−4792.

(7) (a) McDonald, I. M.; Austin, C.; Buck, I. M.; Dunstone, D. J.; Griffin, E.; Harper, E. A.; Hull, R. A. D.; Kalindjian, S. B.; Linney, I. D.; Low, C. M. R.; Pether, M. J.; Spencer, J.; Wright, P. T.; Adatia, T.; Bashall, A. J. Med. Chem. 2006, 49, 2253−2261. (b) McDonald, I. M.; Black, J. W.; Buck, I. M.; Dunstone, D. J.; Griffin, E. P.; Harper, E. A.; Hull, R. A. D.; Kalindjian, S. B.; Lilley, E. J.; Linney, I. D.; Pether, M. J.; Roberts, S. P.; Shaxted, M. E.; Spencer, J.; Steel, K. I. M.; Sykes, D. A.; Walker, M. K.; Watt, G. F.; Wright, L.; Wright, P. T.; Xun, W. J. Med. Chem. 2007, 50, 3101−3112.

(8) Alizadeh, A.; Mokhtari, J. Helv. Chim. Acta 2014, 97, 398−403.

- (9) (a) Rosenström, U.; Sköld, C.; Lindeberg, G.; Botros, M.; Nyberg,
- F.; Karlén, A.; Hallberg, A. J. Med. Chem. 2004, 47, 859–870.
- (b) Rosenström, U.; Sköld, C.; Plouffe, B.; Beaudry, H.; Lindeberg, G.; Botros, M.; Nyberg, F.; Wolf, G.; Karlén, A.; Gallo-Payet, N.; Hallberg, A. J. Med. Chem. 2005, 48, 4009−4024.

(10) Vlasiuk, S. V.; Pavlosky, V. I.; Andronati, S. A.; Gdaniec, M.; Simonov, Y. A. Chem. Heterocycl. Compd. 2000, 36, 1077−1085.

(11) Dong, C.; Xie, L.; Mou, X.; Zhong, Y.; Su, W. Org. Biomol. Chem. 2010, 8, 4827−4830.

(12) Yotphan, S.; Bergman, R. G.; Ellman, J. A. Org. Lett. 2009, 11, 1511−1514.

(13) Bartels, J.; Lu, P.; Maurer, K.; Walker, A. V.; Moeller, K. D. Langmuir 2011, 27, 11199−11205.

(14) Dö rr, A. A.; Lubell, W. D. Heterocycles 2014, 88, 1149−1161.

(15) Crifar, C.; Dörr, A. A.; Lubell, W. D. Tetrahedron Lett. 2015, 56, 3451−3453.

(16) Bourguet, C. B.; Proulx, C.; Klocek, S.; Sabatino, D.; Lubell, W. D. J. Pept. Sci. 2010, 16, 284−296.

(17) Sabatino, D.; Proulx, C.; Klocek, S.; Bourguet, C.; Boeglin, D.; Ong, H.; Lubell, W. D. Org. Lett. 2009, 11, 3650−3653.

(18) Garcia-Ramos, Y.; Proulx, C.; Lubell, W. D. Can. J. Chem. 2012, 90, 985−993.

(19) Tan, S. J.; Xi, H. W.; Bedoura, S.; Lim, K. H. Inorg. Chim. Acta 2012, 384, 29−36.

(20) Burgi, H. B.; Dunitz, J. D.; Shefter, E. J. Am. Chem. Soc. 1973, 95, 5065−5067.

(21) (a) Hein, J. E.; Fokin, V. V. Chem. Soc. Rev. 2010, 39, 1302−1315.

(b) Meldal, M.; Tornøe, C. W. Chem. Rev. 2008, 108, 2952−3015.

(22) Johansson Seechurn, C. C.; Kitching, M. O.; Colacot, T. J.; Snieckus, V. Angew. Chem. Int. Ed. 2012, 51, 5062−5085.

(23) Traoré, M.; Doan, N.; Lubell, W. D. Org. Lett. 2014, 16, 3588− 3591.

(24) De Lucca, G. V.; Otto, M. Bioorg. Med. Chem. Lett. 1992, 2, 1639− 1644.

(25) (a) Pappo, R.; Allen, D. S., Jr.; Lemieux, R. U.; Johnson, W. S. J. Org. Chem. 1956, 21, 478−479. (b) Yu, W.; Mei, Y.; Kang, Y.; Hua, Z.;

Jin, Z. Org. Lett. 2004, 6, 3217−3219.

(26) Bouayad-Gervais, S. H.; Lubell, W. D. Molecules 2013, 18, 14739− 14746.

(27) Rose, G. D.; Gierasch, L. M.; Smith, J. D. Adv. Protein Chem. 1985, 37, 1−109.